

Human Monoclonal Antibody Specifically Binding to
Surface Antigen of Cancer Cell Membrane

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates to a novel human monoclonal antibody useful for diagnosis and therapy of cancer, an isolated DNA encoding the monoclonal antibody, and a hybridoma producing the antibody. The present invention also relates to an anti-cancer formulation comprising the antibody bonded to a liposome which contains an anti-cancer agent.

2. Description of Related Art

There has been no anti-cancer formulation thus far, which is sufficiently effective for the treatment of solid cancer. On the other hand, there has long existed an idea called "targeting" in which a therapeutical agent is concentrated at a tissue or an organ to be treated in order to maximize the therapeutical effect of the agent.

Accordingly, it has been expected that focusing an anti-cancer agent at a cancer tissue by means of "targeting" may allow a therapy of the solid cancer. A number of trials to concentrate an anti-cancer agent or a toxin at a cancer tissue were made since a method for production of mouse monoclonal antibodies in large quantities has been established by Milstein and Köhler (Nature, 1975), and some of them were successful.

Thus far, binding of an antibody to a therapeutic agent has been accomplished by directly binding an antibody to a chemically-modified therapeutic agent, or indirectly binding them via a water-soluble polymer such as dextran.

a 5 These methods, however, have drawbacksⁱⁿ that the amount of a therapeutic agent capable of bindingⁱⁿ to one antibody molecule is very limited, andⁱⁿ that chemical modification of a therapeutic agent often causes lowering of the therapeutical activity. As one of the countermeasures to
10 overcome the drawbacks, there was proposed a new delivery system which consists of an antibody bonded to the surface of a liposome in which a therapeutic agent is encapsuled, and many favorable results were reported (Konno et al, Cancer Research 47 4471, 1987; Hashimoto et al, Japanese
15 Patent Publication (unexamined) No. 134032/1983).

However, mouse monoclonal antibodies have a limited clinical use and continued administration thereof is impossible from a practical point of view due to side effects such as anaphylaxis caused by immune response (See
20 A. Lo Bugli et al, Proc. Natl. Acad. Sci. U.S.A., 86 4220, 1989). Accordingly, human monoclonal antibodies rather than mouse monoclonal antibodies are preferable for the purpose of clinical use. However, preparation of human monoclonal antibodies which adequately react with cancer
25 cells has long been considered very difficult because of

the reasons that it is very difficult to conduct passive immunity for the purpose of obtaining human B cells which produce a desired antibody, and that any efficient methodology which allows infinite reproduction of antibody-producing cells has not been established yet.

In such a situation as mentioned above, the inventors of the present invention have made extensive study for the purpose of obtaining a human monoclonal antibody which permits "targeting therapy" on cancer tissue or organ with the help of anti-cancer agents or toxins, and they have succeeded in preparing a hybridoma capable of producing a novel human monoclonal antibody, the antigen to which exists on the surface of cell membrane of cancer cells. They also have succeeded in preparing a therapeutical formulation useful for "targeting therapy" of cancer, by binding the monoclonal antibody of the invention to a liposome in which an anti-cancer agent is encapsuled.

The present invention is based on these findings.

SUMMARY OF THE INVENTION

Thus, the present invention provides a human monoclonal antibody specific to an antigen existing on the surface of a cancer cell membrane, said monoclonal antibody being produced by a fused cell between a lymphocyte derived from cancer patient and a mouse myeloma cell. The invention further provides an isolated gene encoding the

antibody, a hybridoma producing the antibody, and an anti-cancer formulation containing the antibody.

The human monoclonal antibodies of the present invention contain, in the variable region of the heavy chain, the amino acid sequences shown, for instance, in Sequence Listing Nos. 13, 14, and 15. More specifically, the monoclonal antibodies of the invention include, among others, those in which the variable region of the heavy chain comprises the amino acid sequences shown in Sequence Listing Nos. 16, 17, and 18, and the variable region of the light chain comprises the amino acid sequences shown in Sequence Listing Nos. 19, 20, and 21, and those in which the variable region of the heavy chain comprises the amino acid sequences given in Sequence Listing Nos. 22, 23, and 24, and the variable region of the light chain comprises the amino acid sequences given in Sequence Listing Nos. 25, 26, and 27.

The monoclonal antibodies of the invention include any variants of the above-mentioned specific antibodies, which are obtainable by making insertion, deletion, substitution and/or addition of one or more amino acid residues to the amino acid sequences of the above-identified antibodies with the limitations that such modification must not adversely affect the reactivity of

the antibodies against the antigens. The present invention will be more detailed below.

BRIEF DESCRIPTION OF THE DRAWINGS

~~In the accompanying drawings:~~

Fig. 1 schematically shows the construction of vector pKCRD.

Fig. 2 schematically shows the construction of vector pKCR(ΔE)/H.

Fig. 3 shows reactivity of antibody 1-3-1 to colon cancer cell line C-1.

Fig. 4 shows reactivity of antibody 1-3-1 to gastric cancer cell line MKN45.

Fig. 5 shows anti-cancer effects of adriamycin-containing and PEG-modified liposome bonded to antibody GAH

on the cancer transplanted to nude mouse.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

The hybridoma producing a human monoclonal antibody of the invention is prepared according to the method described by A. Imam (Cancer Research 45 263, 1985). Thus, lymphocytes which have been isolated from extracted lymph node associated with cancer are fused with mouse myeloma cells in the presence of polyethylene glycol. Hybridomas thus obtained are screened by means of enzyme immunoassay using various cancer cell line fixed with paraformaldehyde, and hybridomas capable of producing antibodies are obtained and cultured. From supernatant of the resulting culture, monoclonal antibodies are isolated

and purified according to a conventional method such as disclosed by R. C. Duhamel (J. Immunol. Methods 31 211, 1979).

5 The purified monoclonal antibody is labelled with
a fluorescent substance and examined about its reactivity
with living cancer cells and normal cells such as
erythrocytes and leucocytes using Flow Cytometry.
Hybridoma producing an antibody which reacts with the
living cells but not with normal cells are selected.
10 Alternatively, the reactivity of antibodies to cancer cells
isolated from cancer tissue of a patient is compared with
the reactivity to normal cells derived from non-cancer
segment of the same organ, and a hybridoma producing an
antibody which reacts with the cancer cell and does not
15 react, or reacts as moderately as an antibody derived from
normal volunteer, with normal cells, is selected.

A base sequence of a DNA encoding a human
monoclonal antibody produced by the hybridoma selected
above can be determined in the following manner.

20 In accordance with Casara et al method (DNA 2
329, 1983), mRNAs are separated from the antibody-producing
hybridoma cells, using guanidine thiocyanate-lithium
chloride, and cDNA library is prepared by the use of oligo
(dT) primer. The cDNAs thus obtained are then subjected to
25 (dG) tailing. Consensus sequence between poly C capable of

hybridizing with the dG tail obtained above and an already available human gene encoding heavy or light chain of human antibodies is used as a probe for amplification of the antibody-encoding cDNA by means of PCR. The terminal of the amplified DNA is made blunt. The DNA separated from an electrophoresis gel is inserted to a cloning vector such as pUC119, and the base sequence of the DNA is determined by Sanger et al dideoxy method (Proc. Natl. Acad. Sci. U.S.A. 74 5463, 1977).

Preferable antibodies of the present invention are those in which the variable region of the heavy chain comprises the amino acid sequences shown in Sequence Listing Nos. 13, 14, and 15. Specific examples of preferred antibodies are, among others, those in which the variable region of the heavy chain comprises the amino acid sequences shown in Sequence Listing Nos. 16, 17, and 18, and the variable region of the light chain comprises the amino acid sequences shown in Sequence Listing Nos. 19, 20, and 21, and those in which the variable region of the heavy chain comprises the amino acid sequences shown in Sequence Listing Nos. 22, 23, and 24, and the variable region of the light chain comprises the amino acid sequences shown in Sequence Listing Nos. 25, 26, and 27.

The above-noted amino acid sequences in Sequence Listing Nos. 13, 14, and 15; 16, 17, and 18; and 22, 23,

and 24 are called "hyper variable region" in variable region of the heavy chain. Likewise, the amino acid sequences in Sequence Listing Nos. 19, 20, and 21; 25, 26, and 27 are called "hyper variable region" in variable region of the light chain. These regions are responsible for the specificity of the antibody and determinative to binding affinity between the antibody and the antigenic determinant. Accordingly, the variable region of the heavy chain in the antibodies of the invention can have various amino acid sequences derived from different antibodies so far as it comprises the above-mentioned hyper variable regions.

The most preferred monoclonal antibodies of the invention are those in which the variable regions of the heavy and light chains are represented by the amino acid sequences of Sequence Listing Nos. 5 and 6 respectively, and also 11 and 12 respectively. The DNA sequences encoding constant regions of the heavy and light chains are the same as those disclosed in Nucleic Acids Research 14 1779, 1986, The Journal of Biological Chemistry 257, 1516, 1982 and Cell 22, 197, 1980, respectively.

The monoclonal antibody of the invention may be prepared by culturing the hybridoma producing the antibody of the invention in eRDF or RPMI1640 medium containing fetal bovine serum. Alternatively, it may also be prepared

by connecting the DNAs having the base sequences in
Sequence Listing No. 3, 4, 9 and No. 10, which encode
variable regions of heavy and light chains respectively,
with known DNAs encoding the constant regions as mentioned
5 above to obtain a pair of genes encoding the monoclonal
antibody of the invention, inserting the genes into one of
various known expression vectors, transforming an
appropriate host cell such as CHO cell with the expression
vectors, and culturing the resultant transformant. As
10 expression vectors to be used in animal cells, there may
conveniently used a combination of pKCR (ΔE)/H and pKCRD
which may be constructed in the manner as shown in Figs. 1
and 2 starting from pKCRH2 disclosed by Mishina (Nature 307
605, 1984). In more detail, a gene encoding the heavy
15 chain, to which a HindIII restriction site has been added,
is inserted into plasmid pKCR (ΔE /H) at the HindIII site,
and a selective marker such as DHFR gene is inserted into
the plasmid at SalI site. On the other hand, a gene
encoding the light chain, to both ends of which EcoRI
20 restriction site has been added, is inserted into plasmid
pKCRD at EcoRI site, and then the DHFR gene is also
inserted into the plasmid at SalI site. Both of the
plasmids obtained above are incorporated into a host cell
such as CHO dhfr⁻ (Urlaub G. & Chasin L. A., Proc. Natl.
25 Acad. Sci. U.S.A., 77 4216, 1980) by means of calcium

phosphate method. The resultant transformant is cultured in α MEM medium containing no nucleotide, and grown cells are subjected to further selection for antibody-producing clones. The antibody of the invention can be obtained and
5 purified by culturing the selected clone, adsorbing the resulting supernatant to a column filled with Protein A supported by cerulofine or agarose, and eluting the antibody from the column.

A liposome used for the preparation of the anti-
10 cancer formulation of the invention is composed of two lipid layers. The lipid layer may be of monolayer or multiple layers. Constituents of the liposome are phosphatidylcholine
~~phosphatidylcholine~~, cholesterol, phosphatidylethanolamine, etc. Phosphatidic acid, which provides the liposome with
15 electric charge, may also be added. The amounts of these constituents used for the production of the liposome are, for instance, 0.3-1 mol, preferably 0.4-0.6 mol of cholesterol, 0.01-0.2 mol, preferably 0.02-0.1 mol of phosphatidylethanolamine, 0.0-0.4 mol, preferably 0-0.15mol
20 of phosphatidic acid per 1 mol of phosphatidylcholine.

The liposome used in the present invention may be prepared by conventional methods. For example, a mixture of the above-mentioned lipids, from which the solvents have been removed, is emulsified by the use of a homogenizer,
25 lyophilized, and melted to obtain multilamara liposome.

Adjustment of particle size of the resultant liposomes may be conducted by ultrasonication, high-speed homogenization, or pressure filtration through a membrane having uniform pore size (Hope M. J. et al., *Biochimica et Biophysica Acta* 812 55, 1985). Preferable particle size of the liposomes are between 30nm and 200nm.

Anti-cancer agents encapsuled in the liposome includes carcinostatic agents such as adriamycin, daunomycin, mitomycin, cisplatin, vincristine, epirubicin, methotrexate, 5Fu, and aclacinomycin, toxins such as ricin A and diphtheria toxin, and antisense RNA. Encapsulation of anti-cancer agent into liposome is accomplished by hydration of the lipids with an aqueous solution of the anti-cancer agent. Adriamycin, daunomycin, and epirubicin may be encapsulated into a liposome by means of remote loading method taking advantage of pH gradient (Lawrence D.M. et al., *Cancer Research* 49 5922, 1989).

Binding of a monoclonal antibody to the surface of the liposome mentioned above may be accomplished by the formation of cross-linkage between phosphatidylethanolamine and the antibody using glutaraldehyde. However, preferred method is that a thiolated antibody is allowed to react with a liposome comprising a lipid into which a maleimide group has been incorporated. Remaining maleimide groups on the surface of the liposome may be further reacted with a

compound containing thiolated polyalkyleneglycol moiety,
thereby the surface of the liposome is modified.

a
Thiolation of an ^{antibody} ~~antibody~~ may be conducted by
the use of N-succinimidyl-3-(2-pyridyldithio)propionate
5 (SPDP), which is usually used for thiolation of protein,
iminothiolane, or mercaptoalkylimidate. Alternatively, a
dithiol group intrinsic to an antibody may be reduced to
form a thiol group. The latter is preferred from the view
point of keeping antibody's function. Another method to
10 provide an antibody with a thiol group is that an antibody
is treated with an enzyme such as pepsin to form F(ab)'₂,
which is then reduced with dithiothreitol (DTT) to form
Fab', which gives one to three thiol groups.

The binding of the thiolated antibody to the
15 maleimide group-containing liposome may be accomplished by
reacting them in a neutral buffer solution at pH6.5-7.5 for
2-16 hours.

The anti-cancer formulation of the present
invention may be prepared by means of conventional methods
20 such as dehydration method (Japanese Patent Publication No.
502348/1990) and lyophilization method (Japanese Patent
Publication No. ^{9931/1989} ~~9331/1989~~).

a
The anti-cancer formulation of the invention may
be administered intravascularly, peritoneally, or locally.
25 Dosage of the formulation varies depending on the nature of

particular anti-cancer agent encapsulated into the liposome. When the agent is adriamycin, the dosage is the one corresponding to adriamycin 50mg or less/kg body weight, preferably 10mg or less/kg, more preferably 5mg or less/kg.

The following detailed examples are presented by way of illustration of certain specific embodiments of the present invention.

Example 1

10 Establishment of Hybridoma Producing Human Monoclonal Antibody GAH

Hybridoma producing human monoclonal antibody GAH was established by cell fusion between lymphocytes derived from a lymph node associated with cancer tissue of a patient and mouse myeloma cells.

(1) Preparation of Lymphocytes

Cancer-associated lymph node extracted from a patient suffering from colon cancer was cut up into fine pieces with scissors and scalpel, and cells were dispersed using a stainless net in Culture Medium A (eRDF (Kyokuto Seiyaku Kogyo) + 50µg/ml gentamicin sulfate). The resultant cell suspension was centrifuged at 1000 rpm for 10 minutes and the supernatant was discarded. The residue was suspended in fresh Culture Medium A, and the suspension was centrifuged again to obtain 2.6×10^7 cells.

(2) Cell Fusion

The lymphocyte cells obtained above were subjected to cell fusion with mouse myeloma cells (1×10^7) in the presence of polyethyleneglycol (Boehringer-Mannheim) according to a conventional method. The fused cells were suspended into Culture Medium A added with $10 \mu\text{M}$ hypoxanthine, $0.04 \mu\text{M}$ aminopterin, $1.6 \mu\text{M}$ thymidine, and 10% fetal calf serum (FCS), said medium being referred to as HAT addition medium hereinafter, so that the density of the lymphocytes may be $5.4 \times 10^5/\text{ml}$. The suspension was plated on 96 well plates at $100 \mu\text{l}/\text{well}$ and cultured at 37°C in a CO_2 incubator. Half of the culture medium was substituted with HAT addition medium from time to time and the cultivation was continued until hybridoma's colonies appeared. The hybridoma's colonies were observed in all of the wells. The supernatant of the culture in each well was tested on the reactivity to several established cancer cell lines such as colon cancer cell line C-1 (Sato et al, Igakunoayumi (Progress of Medicine) 96 876, 1976, obtained from Men Eki Seibutsu Kenkyusho (Institute of Immunized Organisms)), and stomach cancer cell line MKN45 (Naito et al, Gan to Kagaku Ryoho (Cancer and Chemotherapy) 5 89, 1978, obtained from above-noted Institute) according to the method described in Experiment 1. Positive wells were 7.3% (35 wells) against C-1 and 4.6% (22 wells) against MKN45,

and 6 wells showed positive reaction to both strains. Cloning of hybridomas was conducted using the wells which showed positive reaction to both lines. The cloning was conducted three times by means of limiting dilution method, and hybridoma clone GAH was established.

Example 2

Purification and Labeling of Monoclonal Antibody GAH

(1) Culture of Hybridoma GAH and Purification of Monoclonal Antibody GAH

Fetal calf serum was passed through a Protein A-agarose (RepliGen), thereby substances adsorbed to the column was removed from the serum. For culture of hybridoma GAH, eRDF culture medium (Kyokuto Seiyaku) to which 3% of the above serum had been added was used. The culture of hybridoma GAH was then charged into a Protein A-agarose column, and adsorbed antibody was then eluted out to obtain purified antibody. The use of the above-noted serum allowed to obtain pure antibody GAH free from other antibodies of serum origin and substances adsorbed to Protein A-agarose. The antibody GAH was confirmed to be a pure IgG by sodium dodecyl sulfate-polyacrylamide gel electrophoresis.

(2) Fluorescent Labeling of Antibody GAH

The purified antibody GAH was labeled by fluorescein isothiocyanate (FITC) according to the method of Coons A. H. Thus, the antibody was dialyzed against a carbonate buffer solution (pH9.5) and reacted with FITC solution. The labeled antibody was separated from free FITC by gel filtration. Absorbance of fractions containing labeled antibody was measured at OD_{280nm} and OD_{495nm} and labeling degree was determined. The binding molar ratio of the antibody and FITC (F/P ratio) was 0.93.

Experiment 1

Study on Reactivity of Human Monoclonal Antibody against Cancer Cell Lines

(1) Cancer Cell Lines and Preservation Thereof

Colon cancer cell line C-1 and stomach cancer cell line MKN45 were used as human cancer cell lines. The cells were preserved and grown at 37°C under 5% CO₂ conditions using Culture Medium B (eRDF medium containing 10% FCS).

(2) Study on Reactivity to Cancer Cell Lines

a. Determination of reactivity against solid cancer cell lines

Cancer cells were cultured until monolayer in a 96 well plate for 3 or 4 days. After removal of culture supernatant, the plate was washed twice by 10mM phosphate buffer (pH7.4) and 0.15M NaCl solution (PBS), and 2% paraformaldehyde fixation was conducted at room temperature

for 20 minutes. After washing 5 times with PBS, PBS solution containing 5% BSA (bovine serum albumin) was added to wells (200 μ /well), and the plate was kept 37°C for 2 hours to complete blocking. The plate was washed 5 times
5 with PBS, and 50 μ l of culture supernatant of hybridoma was added thereto. After two hour reaction at 37°C, the plate was washed 5 times with PBS and 50 μ l of alkaliphosphatase conjugated goat antibody to human antibody (1000 dilution, Capel) was added. Following one hour reaction at 37°C, the
10 plate was washed 5 times with PBS and added with 0.05M carbonate buffer - 1mM MgCl (pH9.5) containing 25mM p-nitrophenyl phosphate at ratio of 50 μ l/well and allowed to react at room temperature for one hour to overnight. Absorbance at 405nm was measured with micro-plate
15 photometer (Colona). Reactivity was determined according to the method described in Example 1 (2). Cloning from the wells in which positive reaction against cultured cancer cell lines C-1 and MKN45 has been observed gave hybridoma GAH. Purified antibody from culture supernatant of GAH
20 showed the same reactivity.

b. Reactivity to living cancer cells

Cancer cells were cultured in a flask or Petri dish and culture supernatant was discarded. To the residue was added a PBS solution containing 0.02% EDTA, and the
25 mixture was left to stand at room temperature for 30

minutes allowing the cells to float. The cells were washed with Culture Medium B by centrifugation and suspended in healthy human serum containing the fluorescent-labeled antibody GAH (final concentration: 50µg/ml) obtained in Example 2 (2) so that cell density of about $1 \times 10^6/200\mu\text{l}$ may be obtained, and the suspension was allowed to react at 0°C for 60 minutes. The suspension was centrifuged at 2000 rpm for 2 minutes and the supernatant was discarded. The remaining cells were suspended in 1ml of PBS, washed by centrifugation, and resuspended in 300µl of PBS containing 10µg/ml of propidium iodide (PI). The suspension was subjected to the observation by flow cytometer (FCM), FACS440 (Becton Dickinson), in order to determine the magnitude of fluorescence (FITC and PI) bonded to particular cell. Dead cells having PI fluorescence could be removed because the dead cells took in PI in the nucleic acids and emitted PI fluorescence. Markers having five standard amounts of fluorescence (quantitative kit: Ortho Diagnostic Systems) were subjected to FCM under the same conditioned as above. Based on the markers, average binding amount of FITC per cell was calculated. On the basis of the average binding amount and F/P ratio of labeled antibody, an average number of antibodies bonded to one living cell was determined. The results are shown in Table 1.

Table 1

Cancer Cell Strain	Antibody	
	GAH	Control IgG
MKN45	3.5×10^4	0.15×10^4
C-1	0.6×10^4	$<0.1 \times 10^4$

When compared with IgG derived from healthy human serum, which was labeled by fluorescence in the same manner as GAH and used as a control, about 6-23 times larger amount of antibody GAH has bonded to stomach and colon cancer cells.

Experiment 2

Reactivity of Human Monoclonal Antibody GAH to Blood Cells

Erythrocytes were separated from peripheral blood taken from 7 healthy volunteers and 3 patients suffering from cancer according to Kinoshita's method (Separation of Erythrocytes; New Edition of Nippon Ketsuekigaku Zensho 13800, 1979).

Leukocytes were obtained in the following manner: Peripheral blood was drawn from healthy volunteers with addition of heparin. 2ml of 6% dextran-physiological saline was added and mixed to 10ml of the blood. The mixture was left to stand at room temperature for 50 minutes to give a plasma layer, which was then separated

and centrifuged at 1500 rpm for 5 minutes to obtain leukocytes.

Reactivities of the monoclonal antibody of the invention to these blood cells were determined by means of FCM in the same manner as in the living cancer cells except that PI was not added. In this connection, the leukocytes were divided into ~~leukocyte~~ ^{lymphocyte} (major leukocyte cell), granulocyte, monocyte, and platelet, based on front and side light scattering in FCM (Bio/Technology 3 337, 1985), and reactivities to respective cells were separately determined. The test results were shown in Table 2.

Table 2

Cells	Antibody	
	GAH	Control IgG
Leukocyte lymphocyte ^{lymphocyte}	negative	negative
granulocyte	0.49×10^4 *	0.48×10^4 *
monocyte	0.41×10^4 *	0.43×10^4 *
platelet	negative	negative
Erythrocyte	negative	negative

*: Average number of antibodies bonded per cell

Antibody GAH showed no reaction to erythrocyte and ~~lymphocyte~~ ^{lymphocyte}, while the reactivity to granulocyte and

monocyte was the same level as the reactivity to control IgG likewise in Experiment 1.

Experiment 3

5

Reactivity of Human Monoclonal Antibody GAH to Cells Derived from Fresh Cancer Tissue and Non-Cancer Tissue

10

In order to study a binding specificity of antibody GAH to cancer cells, normal cells were simultaneously isolated from fresh tissue belonging to the same organ of the same patient from which cancer cells were obtained, and reactivities of antibody GAH to respective cells were determined. Isolation of cells from the tissue was conducted according to Tokita's method (Ganno Rinsho (Cancer in Clinic) 32 1803,1986).

15

Thus, the tissue extracted was placed on Teflon sheet spreaded on a rubber plate, cut with a razor into fine pieces, and transferred onto a 1mm stainless ~~meshes~~ ^{mesh}.

h

h

The ~~meshes~~ ^{mesh} was shaken in a Petri dish full of a culture

20

medium to obtain the medium containing small cell aggregates which passed through the ~~meshes~~ ^{mesh}. The medium was centrifuged at 1000 rpm, and floating fats and suspending necrotic debris were discarded. This centrifugation was repeated several times. The cell aggregates were subjected to pumping by means of a syringe with Cateran needle of 23 gauge to disperse the cells. The reactivity to the cells thus obtained was determined by FCM in the same manner as

u

25

in the living cancer cells. The test results are shown in Table 3.

Table 3

Antibody	Colon		Stomach	
	Cancer Cells	Non-cancer Cells	Cancer Cells	Non-cancer Cells
GAH	1.1×10^4	0.03×10^4	180×10^4	4.6×10^4
Control IgG	0.15×10^4	0.04×10^4	3.5×10^4	0.9×10^4

Average number of antibodies bonded per cell

The average number of GAH antibodies bonded to cancer cells is remarkably higher than that in the non-cancer cells. In addition, the number of antibodies bonded to cancer cells was 51 times greater than that in the control IgG in stomach cancer, and 7 times greater in colon cancer. These results indicate that antibody GAH recognizes an antigen dominantly expressed on the surface of cell membrane of cancer cells.

Examples 3

(1) Determination of Subclass of Light Chain of Monoclonal Antibody GAH

Antibody GAH obtained in Example 2 (1) was subjected to SDS-PAGE in the reduced form. Heavy chain and

light chain separately electrophorated were blotted on a transmembrane (Polyvinylidene-difluoride, Millipore). The membrane was blocked with 5% BSA solution and allowed to react with a goat antibody to human κ or λ chain, which was
5 combined with peroxidase (Capel). After washing, a 0.05% (w/v) 4-chloronaphthol solution containing 0.015% H_2O_2 was allowed to react thereto as a substrate. The light chain of antibody GAH reacted with anti-human κ chain antibody, which was detected through the appearance of ^{Colored}~~coloured~~ band.
10 This revealed that the light chain was κ chain.

(2) Preparation of Gene Encoding Monoclonal Antibody GAH

a. Preparation of cDNA encoding antibody GAH by means of polymerase chain reaction (PCR)

15 According to the method detailed below, poly(A)-containing RNAs were prepared from antibody GAH-producing hybridoma obtained in Example 1 (2) using guanidine thiocyanate-lithium chloride method (DNA 2 329, 1983).

The hybridoma cells (1×10^7) were solubilized in a
20 solution (7.5ml) comprising 5M guanidine thiocyanate, 10mM EDTA, 50mM Tris-HCl, pH7.0, and 8% (v/v) β -mercaptoethanol. To the mixture was further added and mixed cesium chloride to the final concentration of 1g/2.5ml. The solution (8.0ml) was gently overlayed on a 5.7M cesium chloride
25 solution (3.5ml) in a centrifuge tube, and centrifuged at

30,000 rpm for 23.5 hours using Hitachi RPS40T Rotary,
which gave RNAs as a precipitate. The precipitate was
dissolved in a solution (400 μ l) comprising 0.1% sodium
lauryl sulfate, 1mM EDTA, and 10mM Tris-HCl, pH7.5,
5 followed by phenol-chloroform extraction and ethanol
precipitation. The resultant RNAs (about 64 μ g) was
dissolved in a solution (40 μ l) comprising 10mM Tris-HCl,
pH8.0, and 1mM EDTA. A 21 μ l aliquot of the solution
provided about 2.64 μ g of mRNA containing poly(A) by means
of mRNA ~~Purification Kit~~ ^{PURIFICATION KIT} (Pharmacia). The poly(A)-
10 containing mRNA (1.1 μ g) was dissolved in water (10 μ l). To
the solution were added oligo d(T) 12-18 primer (1.5 μ g)
(Pharmacia), 10mM 4 dNTP (3 μ l) (Takara Shuzo), reverse
transcriptase (40U) (Life Science), RNase inhibitor (30U)
15 (Takara Shuzo), 5 \times reverse transcriptase buffer (6 μ l)
comprising 250mM Tris-HCl, pH8.3, 40mM magnesium chloride,
and 250mM potassium chloride, and additionally water to
make a total volume of 30 μ l. The mixture was allowed to
react at 41°C for one hour, followed by ethanol
20 precipitation to obtain cDNA.

The cDNA thus obtained was dissolved in water
(15.3 μ l). To the solution were added a 5 \times terminal
deoxynucleotide transferase buffer (4.8 μ l) (250mM Tris-HCl,
pH7.5, 50mM magnesium chloride), terminal deoxynucleotide
25 transferase (12U) (Pharmacia), and 10mM dGTP (2.4 μ l)

(Takara Shuzo) to make a total volume of 24 μ l, and the mixture was allowed to react at 37°C for 1.5 hours to add poly d(G) at 3' terminal of cDNA. After completion of the reaction, the enzymes were inactivated by heating at 70°C for 15 minutes.

a
PCR was conducted based on the cDNA thus obtained as a template using Perkin Elmer Cetus DNA ~~Thermal Cycler~~ ^{THERMAL CYCLER (PCR apparatus)} following the manual provided by the manufacturer. Thus, to the above reaction mixture (2 μ l) were added, as a primer for amplifying cDNA encoding variable region of the heavy chain, poly C (15 nucleotides) which hybridizes dG tail added to 3' terminal of the cDNA (40pmol), a single stranded DNA primer (37 nucleotides) corresponding to the region spanning from part of the variable region (113-119 amino acid sequence in Sequence Listing No. 5) to the constant region which is common to all human IgGs, (25pmol) (Nucleic Acids Research 14 1779, 1986), poly C as a primer for amplifying cDNA encoding variable region of the light chain (40pmol), a single stranded DNA primer (21 nucleotides) corresponding to the region spanning from J region of human κ chain (113-114 amino acid sequence of Sequence Listing No. 6) to the constant region, (The Journal of Biological Chemistry 257 1516, 1982; Cell 22 197, 1980) (40pmol), 10 \times PCR buffer (100mM Tris-HCl, pH8.3, 500mM potassium chloride, 15mM magnesium chloride, 0.1% (w/v)

10
15
20
25

a
n

Shown in Sequence Listing No. 1
Shown in Sequence Listing No. 2

gelatin (10 μ l), 10mM 4 dNTP (2 μ l) (Takara Shuzo), and Taq DNA polymerase (2.5U) (Takara Shuzo)), and further water to make a final volume of 100 μ l. Thirty cycles of incubations at 94°C for one minute (denaturing step) at 55°C for two minutes (annealing step) and at 72°C for three minutes (elongation step) were conducted and further incubation at 72°C for seven minutes was added. Reaction mixture was subjected to ethanol precipitation, and resultant precipitates were dissolved in water (30 μ l).

To the aqueous solution were added Klenow fragment (2U) (Takara Shuzo), 1mM 4 dNTP (4 μ l), and 10 \times blunting buffer (500mM Tris-HCl, pH7.6, 100mM magnesium chloride) (4 μ l), 40 μ l in total, and the mixture was allowed to react at 37°C for 30 minutes to obtain a double-stranded cDNA having blunt ends.

b. Determination of base sequence of cDNA

The cDNA solution obtained above was subjected to 2% agarose electrophoresis, and a band was observed at about 500bp. The band was cut away from the agarose gel. The cDNA was inserted into a cloning vector pUC119 at SmaI site, and the base sequence was determined by dideoxy method, which revealed that among total base sequence of the PCR fragment, the base sequences encoding variable regions of the heavy and light chains were respectively those shown in Sequence Listing Nos. 3 and 4.

The amino acid sequences of variable regions of heavy and light chains of antibody GAH produced by the above-mentioned hybridoma were deduced from the base sequences determined above and are respectively shown in Sequence Listing Nos. 5 and 6. Based on the base sequences determined, antibody GAH was shown to belong to IgG1 subclass. The DNA fragment, the base sequence of which has been determined, can be prepared by means of DNA synthesizer with good reproducibility, and therefore, the acquisition of the DNA fragment does not require the repetition of the above procedure.

Example 4

Establishment of Human Monoclonal Antibody 1-3-1 Producing Hybridoma by Cell Fusion between Lymphocyte Derived from Cancer Associated Lymph Node and Mouse Myeloma

(1) Preparation of Lymphocyte

In substantial accordance with the procedure described in Example 1 (1), lymphocytes (3×10^7) were prepared starting from cancer associated lymph node extracted from a patient with lung cancer.

(2) Cell Fusion

Lymphocyte cells obtained above were fused with mouse myeloma cells (8×10^6) using polyethyleneglycol (Boehringer-Mannheim) according to the conventional method. In the same manner as Example 1 (2), the fused cells were

suspended in HAT addition medium to obtain cell density of $5.2 \times 10^5/\text{ml}$ and placed on a 96 well plate at a ratio of 100 μl /plate. Half of the culture medium was substituted with HAT addition medium from time to time and the culture was continued until hybridoma's colonies appeared. The hybridoma's colonies were observed in all of the wells. In the same manner as in Example 1 (2), the supernatant of the culture in each well was tested on the reactivity to fixed cancer cell lines such as colon cancer cell line C-1 and stomach cancer cell line MKN45, in accordance with the procedure described in Experiment 1 (2)-a. Positive wells were 16.3% (94 well) against C-1 and 6.3% (36 wells) against MKN45, and 4 wells showed positive reaction to both lines.

Cloning of hybridoma cells was conducted using the wells which showed positive reaction to both lines. The cloning was conducted three times by means of limiting dilution method, and hybridoma clone 1-3-1 was established.

Example 5

Purification and Labeling of Monoclonal Antibody 1-3-1

(1) Culture of Hybridoma 1-3-1 and Purification of Monoclonal Antibody 1-3-1

For culture of hybridoma 1-3-1, eRDF culture medium (Gokuto Seiyaku) to which 3% of the serum described

in Example 2 (1) had been added was used. The culture of hybridoma 1-3-1 was then charged into a Protein A-agarose column, and adsorbed antibody was then eluted out to obtain purified antibody 1-3-1. The antibody was confirmed to be a pure IgM by SDS-PAGE.

(2) Fluorescent Labeling of Antibody 1-3-1

The purified antibody 1-3-1 was labeled by FITC according to the method described in Example 2 (2). Absorbance of fractions containing labeled antibody was measured at OD_{280nm} and OD_{495nm}, and labeling degree was determined. F/P ratio was 6.7.

Experiment 4

Study on Reactivity of Human Monoclonal Antibody to Cancer Cell Lines

(1) Cancer Cell Lines and Preservation thereof

Human colon cancer cell line C-1 and stomach cancer cell line MKN45 were preserved and grown at 37°C and 5% CO₂ conditions in Culture Medium B in the same manner as described in Experiment 1 (1).

(2) Study on Reactivity to Living Cancer Cell Lines

Cancer cells were cultured in a flask or Petri dish and culture supernatant was discarded. To the residue was added PBS solution containing 0.02% EDTA, and the mixture was left to stand at room temperature for 30

minutes allowing the cells to float. The cells were washed with Culture Medium B by centrifugation and suspended in PBS so as to make the cell density of about $1 \times 10^6/200\mu\text{l}$. Antibody 1-3-1 obtained in Example 5 (1) was added to the
5 above solution to make the final concentration of the antibody of $50\mu\text{g/ml}$, and the mixture was allowed to react at 0°C for 60 minutes. The suspension was centrifuged at 2000 rpm for 2 minutes and the supernatant was discarded. To the remaining cells was added FITC labeled anti-human
10 antibody solution ($200\mu\text{l}$) (Capel) diluted with 1% BSA-containing PBS by 500 times, and the resulting cell suspension was kept at 0°C for 60 minutes. The suspension was centrifuged at 2000 rpm for 2 minutes to remove the supernatant, and the remaining cells was suspended in and
15 washed with PBS (1ml) by centrifugation, and the cells were finally suspended in PBS ($300\mu\text{l}$) containing PI ($10\mu\text{g/ml}$). The resultant cell suspension was subjected to FCM, and magnitude of fluorescence (FITC and PI) bonded to particular cell was determined. The reactivities of
20 antibody 1-3-1 to colon cancer cell line C-1 and stomach cancer cell line MKN45 are respectively shown in Figs. 3 and 4 of the accompanying drawings. In the figures, the abscissa shows fluorescence intensity per cancer cell and the ordinate shows the number of the cancer cells. As a
25 control, a commercially available IgM antibody (Capel) was

used, and the reactivities of the IgM antibody to the above-identified cancer cells were determined. In the figures, the dotted line and solid line show the reactivities of antibody 1-3-1 and the control respectively. These figures show that antibody 1-3-1 has a strong binding ability to cancer cells.

Experiment 5

Reactivity of Human Monoclonal Antibody 1-3-1 to Cells Derived from Fresh Cancer Tissue and Non-Cancer Tissue

In order to study a binding specificity of antibody 1-3-1 to cancer cells, normal cells were simultaneously isolated from fresh tissue belonging to the same organ of the same patient, from which cancer cells were obtained, and reactivities of antibody 1-3-1 to respective cells were measured. Isolation of cells from the tissue was conducted according to Tokita's method as described in Experiment 3.

The reactivity to the cells obtained above was determined by FCM in the same manner as previously described in the living cancer cells. However, the cells were washed with Culture Medium B, suspended in serum derived from healthy volunteers, which serum contained fluorescent labeled antibody 1-3-1 (final concentration of 50 μ g/ml) prepared in Example 5 (2), to the cell density of about $1 \times 10^6 / 200 \mu$ l. The suspension was allowed to react at

0°C for 60 minutes and subjected to centrifugation at 2000 rpm for 2 minutes to remove the supernatant. The remaining cells were suspended in PBS (1ml) and washed by centrifugation. The cells were resuspended in PBS (300µl) containing PI (10µg/ml), and the suspension was subjected to FCM. The amount of fluorescent (FITC and PI) bonded to a particular cell was measured. Markers (5 species) for determining the amount of fluorescence (quantitative kit as previously prescribed) were subjected to FCM under the same condition. Average amount of FITC bonded to a single cell was calculated. Based on the average amount and F/P ratio of labeled antibody calculated in Example 5 (2), the average number of antibodies bonded to a living cancer cell was calculated. The results are shown in Table 4.

Table 4

Antibody	Colon		Stomach	
	Cancer Cells	Non-cancer Cells	Cancer Cells	Non-cancer Cells
1-3-1	1.5×10^4	0.04×10^4	1.8×10^3	0.05×10^3
Control	0.15×10^4	0.04×10^4	0.2×10^3	0.3×10^3

The reactivity of the human monoclonal antibody 1-3-1 to non-cancer cells was the same level as, or less than, that of the antibody which was derived from

peripheral blood of healthy volunteers and fluorescent-labeled in the same manner as antibody 1-3-1, while the average number of antibodies bonded to cancer cells is remarkably higher than that in the non-cancer cells. In addition, the number of antibodies bonded to cancer cells was 10 times greater than that in the control antibody both in stomach and colon cancer. These results indicate that antibody 1-3-1 recognizes an antigen dominantly expressed on the surface of cell membrane of cancer cells.

Examples 6

(1) Determination of Subclass of Light Chain of Monoclonal Antibody 1-3-1

In order to determine the subclass of the light chain of antibody 1-3-1, the same procedure as described in Example 3 was repeated except that antibody 1-3-1 obtained in Example 5 (1) was used in place of antibody GAH. The light chain of antibody 1-3-1 reacted with anti-human λ chain antibody, which was detected through the appearance of coloured band. This revealed that the light chain was λ chain.

(2) Preparation of Gene Encoding Monoclonal Antibody 1-3-1 and Determination of Base Sequence

a. Preparation of cDNA encoding antibody 1-3-1 by means of PCR

According to the method detailed below, poly(A) containing RNAs were prepared from antibody 1-3-1 producing

hybridoma obtained in Example 4 (2) using guanidine thiocyanate-lithium chloride method (DNA 2 329, 1983).

In the same manner as described in Example 3 except that the number of hybridoma cells used was 2×10^8 ,
5 the mRNA was prepared. The resultant RNAs (about 1.8mg) was dissolved in a solution (1ml) comprising 10mM Tris-HCl, pH8.0, and 1mM EDTA. A 230 μ l aliquot of the solution provided about 20 μ g of mRNA containing poly(A) after
A PURIFICATION KIT Kit for Purifying mRNA Med purification by means of mRNA ~~Purification Kit~~ (Pharmacia).
10 Following the procedure described in Example 3, the poly(A)-containing mRNA (4.3 μ g) was dissolved in water (10 μ l), and to the solution were added oligo d(T) 12-18 primer (0.6 μ g), 10mM 4 dNTP (2 μ l), reverse transcriptase (40U), RNase inhibitor (30U), 5 x reverse transcriptase
15 buffer (6 μ l), and additionally water to make a total volume of 30 μ l. The mixture was allowed to react at 42°C for one hour, followed by ethanol precipitation to obtain cDNA.

The cDNA thus obtained was dissolved in water (20 μ l). To the solution were added a 5 x terminal
20 deoxynucleotide transferase buffer (5 μ l), terminal deoxynucleotide transferase (11U), and 10mM dGTP (2.5 μ l) to make a total volume of 25 μ l by adding water (6.5 μ l), and the mixture was allowed to react at 37°C for 1 hour to add poly d(G) at 3' terminal of cDNA. After completion of the

reaction, the enzymes were inactivated by heating at 70°C for 10 minutes.

PCR was conducted using the cDNA thus obtained as a template. Thus, to the above reaction mixture (2.5µl) were added, as a primer for amplifying cDNA encoding variable region of the heavy chain, poly C (14 nucleotides) which hybridizes dG tail added to 3' terminal of the cDNA (25pmol), a single stranded DNA primer (17 nucleotides) corresponding to the base sequence of constant region of IgM shown in Sequence Listing No. 7 (25pmol) (Nucleic Acids Research 18 4278, 1990), poly C as a primer for amplifying cDNA encoding variable region of the light chain (25pmol), a single stranded DNA primer (19 nucleotides) (25pmol) corresponding to the base sequence of constant region of λ chain, shown in Sequence Listing No. 8 (Nature 294 536, 1981). The mixture was treated in the same manner as described in Example 3, which provided a double-stranded cDNA having blunt ends.

b. Determination of base sequence of cDNA

The cDNA solution obtained above was subjected to 2% agarose electrophoresis, and a band was observed at about 500bp. The band was cut away from the agarose gel. The cDNA was inserted into a cloning vector pUC119 at SmaI site, and the base sequence was determined by dideoxy method, which revealed that among total base sequence of

the PCR fragment, the base sequence encoding variable regions of the heavy and light chains were respectively those shown in Sequence Listing Nos. 9 and 10.

The amino acid sequences of variable regions of heavy and light chains of antibody 1-3-1 produced by the above-mentioned hybridoma were deduced from the base sequences determined above and are respectively shown in Sequence Listing Nos. 11 and 12. The DNA fragment, the base sequence of which has been determined, can be prepared by means of DNA synthesizer with good reproducibility, and therefore, the acquisition of the DNA fragment does not require the repetition of the above procedure.

Example 7

Preparation of Adriamycin-Containing Liposome Bonded to Antibody GAH

a. Preparation of Thiolated Antibody

Anti-cancer antibody GAH (IgG) was dissolved in 0.1M - acetate buffer (pH4.0), and pepsin (1/40 mol) (Cooper Biomedical) was added thereto. The mixture was allowed to react overnight to prepare $F(ab')_2$. Chromatography over cation-exchange resin (mono S) (Pharmacia) isolated $F(ab')_2$. The solvent used was a linear gradient of 0.1M - acetate buffer (pH4.0) containing 0-0.5M NaCl. To the isolated $F(ab')_2$ in 0.1M - acetate buffer (pH4.5) containing 0.15M NaCl was added DTT at a ratio of

12 μ l of 10% DTT/1mg antibody. The mixture was left to stand at room temperature for 80 minutes. After completion of reaction, the mixture was passed through a gel filtration column (PD-10) equilibrated with PBS for desalification to obtain thiolated Fab'.

b. Thiolation of polyethylene/glycol

L-cysteine (48mg) was dissolved in 0.4M borate buffer (10ml), and 2,4-bis (polyethylene glycol)-6-chloro-s-triazine (200mg) (activated PEG 2) (Seikagaku Kogyo) was added thereto. The mixture was allowed to react at room temperature overnight. To the resultant PEG bonded with cysteine was added DTT (62mg), and the mixture was allowed to react at 37°C for 6 hours to obtain a solution containing PEG bonded with cysteine. The solution was gel filtered (GH-25, Seikagaku Kogyo) for desalting, and the solvent was substituted by 10mM phosphate buffer (pH7.4) and 0.15M-NaCl (PBS). The solution was added to thiopropyl Sepharose 6B (Pharmacia) equilibrated with PBS, and non-bonded substances were washed away by PBS. Cysteine-binding PEG adsorbed to the gel was eluted out by PBS containing 50mM-DTT, which was then subjected to gel filtration to remove excessive DTT. This gave thiolated PEG.

c. Maleimidation of dipalmitoylphosphatidyl-ethanolamine

Dipalmitoylphosphatidylethanolamine (127mg), N-(ϵ -maleimidecaproyloxy)succinimide (EMCS) (80mg), and triethylamine were added to a chloroform/methanol (5:1) solution (44 μ l), and the mixture was allowed to react for 3
5 hours under nitrogen gas. Additional EMCS (20mg) was added and the mixture was allowed to react at room temperature for further 3 hours. After confirmation of negative ninhydrin reaction of the reaction mixture, the mixture was evaporated to dryness under reduced pressure and the
10 residue was dissolved in a trace amount of chloroform. The maleimidated dipalmitoylphosphatidylethanolamine thus obtained was purified by chromatography over UNISIL (Gasukuro Kogyo) equilibrated with chloroform, using a chloroform/methanol (10:1) solution as an eluent.

15 d. Preparation of liposome containing adriamycin bearing maleimide group

Solid lipid mixture (100mg) (Nippon Seika), which consists of dipalmitoylphosphatidylcholine (DPPC), cholesterol (Chol), and maleimidated
20 dipalmitoylphosphatidylethanolamine at a ratio of 18:10:0.5 (mol) was added to 0.3M citrate buffer (pH4) (1ml) and admixed. Freezing and thawing of the mixture was repeated 5 times to achieve hydration. This gave multimeric liposome. The liposome was charged in an extruder (Lipe x
25 Biomembranes) equipped with a polycarbonate membrane

(Nucleopore; Microscience) having a pore size of 200nm and kept at 60°C. Repeated pressure-filtration (10 times) gave a dressed liposome. The liposome solution was neutralized with addition of 1M NaOH solution, and to the neutral solution was added one tenth (by weight) of adriamycin (Kyowa Hakko) with respect to the lipid components while being kept at 60°C. More than 97% of adriamycin was positively enclosed into the liposome according to the pH slope between the inside and outside of the liposome to give a liposome into which adriamycin bearing maleimide group had been encapsulated.

e. Binding of maleimide group-bearing adriamycin-encapsulated liposome to thiolated antibody and PEG modification

To the adriamycin-encapsulated liposome obtained above (lipid components: 100mg) was added thiolated Fab' antibody (5mg), and the mixture was allowed to react at 37°C for 8 hours. To the reaction mixture was added thiolated PEG (5μmol), and the mixture was allowed to react in PBS at room temperature for 6 hours to obtain adriamycin-encapsulated liposome bonded to antibody and modified with PEG. The latter was further subjected to gel filtration using ^{SEPHAROSE} Sepharose C16B (Pharmacia) to remove non-reacted cysteine-binding PEG.

Confirmation of Pharmaceutical Effectiveness of
Adriamycin-Encapsulated Liposome Bonded to
Antibody GAH and Modified with PEG

Study on anti-cancer effect of antibody GAH was
5 conducted in the manner as described below using human
stomach cancer cell line MKN45 which had shown reactivity
to antibody GAH and accumulative behavior in
transplantation to nude mouse.

Cultured MKN45 cells (1×10^6) were subcutaneous-
10 transplanted to nude mouse. Experiment started when the
cancer weight became about 100 μ g after ten days from the
transplantation (Fig. 5). Adriamycin-encapsulated liposome
bonded to antibody GAH and modified with PEG was
administered to mouse via caudal vein at a dose
15 corresponding to 5mg/kg or adriamycin day 0, 3, 7 (shown by
mark \diamond in Fig. 5). As a control, phosphate buffered
physiological saline (shown by mark \blacklozenge), adriamycin (shown
by mark \square), and adriamycin-encapsulated liposome modified
with PEG (shown by mark \times) were administered to mice (each
20 6-7 animals). Time-course measurement of growth of cancer
was conducted by means of Battle-Columbus method wherein
presumptive cancer weight was determined according to the
formulation : (short diameter) \times (short diameter) \times (long
diameter)/2, and compared with that determined at the
25 beginning of the experiment.

In Fig. 5, the abscissa shows time-lapse (days) after beginning of the experiment, and the mark (↓) indicates the administration of the pharmaceutical formulation of the invention. Fig. 5 clearly shows that the formulation of the invention, adriamycin-encapsulated liposome bonded to antibody GAH, possesses potent anti-cancer effect. It is apparent, therefore, that human monoclonal antibodies of the invention allow continuous and long term "targeting therapy" of cancer tissue or organ with the help of anti-cancer agents or toxins.

(1) GENERAL INFORMATION:

(i) APPLICANT: Saiko HOSOKAWA
Toshiaki TAGAWA
Yoko HIRAKAWA
Norihiko ITO
Kazuhiro NAGAIKE

(ii) TITLE OF INVENTION: Human Monoclonal Antibody
Specifically Binding to Surface
Antigen of Cancer Cell Membrane

(iii) NUMBER OF SEQUENCES: 42

(2) INFORMATION FOR SEQ ID NO:1

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE: human IgG antibody

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

G GCC CTT GGT GGA GGC TGA AGA GAC GGT GAC CAT TCT

37

(2) INFORMATION FOR SEQ ID NO:2

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE: human IgG antibody

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TGG TGC AGC CAC AGT TCG TTT

21

(2) INFORMATION FOR SEQ ID NO:3

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 357 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CAG GTG CAG CTG CAG GAG TCG GGC CCA GGA CTG GTG AAG CCT TCA	45
CAG ACC CTG TCC CTC ACC TGC ACT GTC TCT GGT GGC TCC ATC AGC	90
AGT TGT GGT TTC TAC TGG AAC TGG ATC CGC CAG CAC CCA GGG AAG	135
GGC CTG GAG TGG ATT GGG TAC ATC TAT TAC AGT GGG AGC ACC TAC	180
TAC AAC CCG TCC CTC AAG AGT CGA GTT ACC ATA TCG CTA GAC ACG	225
TCT AAG AGC CAG TTC TCC CTG AAG CTG AGC TCT CTG ACT GCC GCG	270
GAC ACG GCC GTG TAT TAC TGT GCG AGG TCT ACC CGA CTA CGG GGG	315
GCT GAC TAC TGG GGC CAG GGA ACA ATG GTC ACC GTC TCT TCA	357

(2) INFORMATION FOR SEQ ID NO:4

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 342 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GAC ATC GTG ATG ACC CAG TCT CCA GAC TCC CTG GCT GTG TCT CTG	45
GGC GAG AGG GCC ACC ATC AAC TGC AAG TCC AGC CAG AGT GTT TTA	90
TAC AAC TCC AAC AAT AAG AAA TAC TTA GCT TGG TAC CAG CAG AAA	135
CCA GGA CAG CCT CCT AAG CTG CTC ATT TAC TGG GCA TCT ACC CGG	180

10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 114 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Asp	Ile	Val	Met	Thr	Gln	Ser	Pro	Asp	Ser	Leu	Ala	Val	Ser	Leu
1				5					10					15
Gly	Glu	Arg	Ala	Thr	Ile	Asn	Cys	Lys	Ser	Ser	Gln	Ser	Val	Leu
				20					25					30
Tyr	Asn	Ser	Asn	Asn	Lys	Lys	Tyr	Leu	Ala	Trp	Tyr	Gln	Gln	Lys
				35					40					45
Pro	Gly	Gln	Pro	Pro	Lys	Leu	Leu	Ile	Tyr	Trp	Ala	Ser	Thr	Arg
				50					55					60
Glu	Ser	Gly	Val	Pro	Asp	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Gly	Thr
				65					70					75
Asp	Phe	Thr	Leu	Thr	Ile	Ser	Ser	Leu	Gln	Ala	Glu	Asp	Val	Ala
				80					85					90
Val	Tyr	Tyr	Cys	Gln	Gln	Tyr	Tyr	Ser	Thr	Pro	Trp	Thr	Phe	Gly
				95					100					105
Gln	Gly	Thr	Lys	Val	Glu	Ile	Lys	Arg						
				110					114					

(2) INFORMATION FOR SEQ ID NO:7

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE: human IgM antibody

TAC TAC GGT ATG GAC GTC TGG GGC CAA GGG ACC ACG GTC ACC GTC	360
TCC TCA	366

(2) INFORMATION FOR SEQ ID NO:10

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 324 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TAT GAG CTG ACA CAG CCA CCC TCG GTG TCA GTG TCC CCA GGA CAG	45
ACG GCC AGG ATC ACC TGC TCT GGA GAT GCA TTG CCA AAG CAA TAT	90
GCT TAT TGG TAC CAG CAG AAG CCA GGC CAG GCC CCT GTG CTG GTG	135
ATA TAT AAA GAC AGT GAG AGG CCC TCA GGG ATC CCT GAG CGA TTC	180
TCT GGC TCC AGC TCA GGG ACA ACA GTC ACG TTG ACC ATC AGT GGA	225
GTC CAG GCA GAA GAC GAG GCT GAC TAT TAC TGT CAA TCA GCA GAC	270
AGC AGT GGT ACT TAT GAG GTA TTC GGC GGA GGG ACC AAG CTG ACC	315
GTC CTA GGT	324

(2) INFORMATION FOR SEQ ID NO:11

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 122 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

- 48 -

Gln	Leu	Gln	Leu	Gln	Glu	Ser	Gly	Pro	Gly	Leu	Val	Lys	Pro	Ser
1				5					10					15
Glu	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Gly	Ser	Ile	Ser
				20					25					30
Ser	Ser	Ser	Tyr	Tyr	Trp	Gly	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys
				35					40					45
Gly	Leu	Glu	Trp	Ile	Gly	Ser	Ile	Tyr	Tyr	Ser	Gly	Ser	Thr	Tyr
				50					55					60
Tyr	Asn	Pro	Ser	Leu	Lys	Ser	Arg	Val	Thr	Ile	Ser	Val	Asp	Thr
				65					70					75
Ser	Lys	Asn	Gln	Phe	Ser	Leu	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala
				80					85					90
Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala	Arg	Gly	Ser	Tyr	Gly	Gly	Tyr
				95					100					105
Tyr	Tyr	Gly	Met	Asp	Val	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val
				110					115					120
Ser	Ser													
				122										

(2) INFORMATION FOR SEQ ID NO:12

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 108 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Tyr	Glu	Leu	Thr	Gln	Pro	Pro	Ser	Val	Ser	Val	Ser	Pro	Gly	Gln
1				5					10					15
Thr	Ala	Arg	Ile	Thr	Cys	Ser	Gly	Asp	Ala	Leu	Pro	Lys	Gln	Tyr

	20		25		30
Ala	Tyr	Trp	Tyr	Gln	Gln
			Lys	Pro	Gly
				Gln	Ala
				Pro	Val
				Leu	Val
	35		40		45
Ile	Tyr	Lys	Asp	Ser	Glu
			Arg	Pro	Ser
				Gly	Ile
				Pro	Glu
				Arg	Phe
	50		55		60
Ser	Gly	Ser	Ser	Ser	Gly
			Thr	Thr	Val
				Thr	Leu
				Thr	Ile
				Ser	Gly
	65		70		75
Val	Gln	Ala	Glu	Asp	Glu
			Ala	Asp	Tyr
				Tyr	Cys
				Gln	Ser
				Ala	Asp
	80		85		90
Ser	Ser	Gly	Thr	Tyr	Glu
			Val	Phe	Gly
				Gly	Gly
				Gly	Thr
				Lys	Leu
				Thr	
	95		100		105
Val	Leu	Gly			
	108				

(2) INFORMATION FOR SEQ ID NO:13

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 8 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: hybridoma producing human monoclonal antibody, an antigen to which exists on the surface of cancer cell membrane

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Ile Ser Ser Xaa Xab Xac Tyr Trp

1

5

Xaa : Cys or Ser, Xab : Gly or Ser, Xac : Phe or Tyr

(2) INFORMATION FOR SEQ ID NO:14

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 12 amino acids

- 50 -

(B) TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: protein
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: hybridoma producing human monoclonal antibody, an antigen to which exists on the surface of cancer cell membrane

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Ile Gly Xaa Ile Tyr Tyr Ser Gly Ser Thr Tyr Tyr
1 5 10
Xaa : Tyr or Ser,

(2) INFORMATION FOR SEQ ID NO:15

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: protein
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: hybridoma producing human monoclonal antibody, an antigen to which exists on the surface of cancer cell membrane

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Gly Xaa Asp Xab
1

Xaa : Ala or Met, Xab : Tyr or Val

(2) INFORMATION FOR SEQ ID NO:16

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 amino acids
(B) TYPE: amino acid

- 51 -

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (vi) ORIGINAL SOURCE:
 - (G) CELL TYPE: Hybridoma producing human antibody GAH
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Ile Ser Ser Cys Gly Phe Tyr Trp Asn
1 5

- (2) INFORMATION FOR SEQ ID NO:17
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (vi) ORIGINAL SOURCE:
 - (G) CELL TYPE: Hybridoma producing human antibody GAH
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Ile Gly Tyr Ile Tyr Tyr Ser Gly Ser Thr Tyr Tyr
1 5 10

- (2) INFORMATION FOR SEQ ID NO:18
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (vi) ORIGINAL SOURCE:
 - (G) CELL TYPE: Hybridoma producing human antibody GAH
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Ser Thr Arg Leu Arg Gly Ala Asp Tyr

1

5

(2) INFORMATION FOR SEQ ID NO:19

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Lys	Ser	Ser	Gln	Ser	Val	Leu	Tyr	Asn	Ser	Asn	Asn	Lys	Lys	Tyr	Leu	Ala
1					5					10					15	

(2) INFORMATION FOR SEQ ID NO:20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 7 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Trp	Ala	Ser	Thr	Arg	Glu	Ser
1					5	

(2) INFORMATION FOR SEQ ID NO:21

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 9 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Gln Gln Tyr Tyr Ser Thr Pro Trp Thr
1 5

(2) INFORMATION FOR SEQ ID NO:22

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ile Ser Ser Ser Ser Tyr Tyr Trp Gly Trp
1 5 10

(2) INFORMATION FOR SEQ ID NO:23

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 14 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

- 54 -

Ile Gly Ser Ile Tyr Tyr Ser Gly Ser Thr Tyr Tyr Asn Pro
1 5 10

(2) INFORMATION FOR SEQ ID NO:24

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 12 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Gly Ser Tyr Gly Gly Tyr Tyr Tyr Gly Met Asp Val
1 5 10

(2) INFORMATION FOR SEQ ID NO:25

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 9 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Asp Ala Leu Pro Lys Gln Tyr Ala Tyr
1 5

(2) INFORMATION FOR SEQ ID NO:26

(i) SEQUENCE CHARACTERISTICS:

- 55 -

(A) LENGTH: 4 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: protein
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Lys Asp Ser Glu

1

(2) INFORMATION FOR SEQ ID NO:27

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 11 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: protein
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Gln Ser Ala Asp Ser Ser Gly Thr Tyr Glu Val

1

5

10

(2) INFORMATION FOR SEQ ID NO:28

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: cDNA
(vi) ORIGINAL SOURCE:

- 56 -

(G) CELL TYPE: hybridoma producing human monoclonal antibody, an antigen to which exists on the surface of cancer cell membrane

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

ATC AGC AGT WGT RGT TWC TAC TGG 28
W : T or A, R : G or A

(2) INFORMATION FOR SEQ ID NO:29

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: hybridoma producing human monoclonal antibody, an antigen to which exists on the surface of cancer cell membrane

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

ATT GGG WRY ATC TAT TAY AGT GGG AGC ACC TAC TAC 36
W : T or A, R : A or G, Y : C or T

(2) INFORMATION FOR SEQ ID NO:30

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: hybridoma producing human monoclonal antibody, an antigen to which exists on the surface of cancer cell membrane

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGK RYK GAC KWC 12
K : G or T, R : G or A, Y : C or T
W : A or T

(2) INFORMATION FOR SEQ ID NO:31

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

ATC AGC AGT TGT GGT TTC TAC TGG 27

(2) INFORMATION FOR SEQ ID NO:32

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

ATT GGG TAC ATC TAT TAC AGT GGG AGC ACC TAC TAC 36

(2) INFORMATION FOR SEQ ID NO:33

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs

- 58 -

(B) TYPE: nucleic acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: cDNA
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: Hybridoma producing human antibody GAH
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

TCT ACC CGA CTA CGG GGG GCT GAC TAC 27

(2) INFORMATION FOR SEQ ID NO:34

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 51 base pairs
(B) TYPE: nucleic acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: cDNA
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: Hybridoma producing human antibody GAH
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

AAG TCC AGC CAG AGT GTT TTA TAC AAC TCC AAC AAT AAG AAA TAC TTA GCT
51

(2) INFORMATION FOR SEQ ID NO:35

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: cDNA
(vi) ORIGINAL SOURCE:
(G) CELL TYPE: Hybridoma producing human antibody GAH
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

TGG GCA TCT ACC CGG GAA TCC 21

(2) INFORMATION FOR SEQ ID NO:36

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody GAH

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

CAG CAG TAT TAT AGT ACT CCG TGG ACG 27

(2) INFORMATION FOR SEQ ID NO:37

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

ATC AGC AGT AGT AGT TAC TAC TGG GGC TGG 30

(2) INFORMATION FOR SEQ ID NO:38

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 42 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

- 60 -

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

ATT GGG AGT ATC TAT TAT AGT GGG AGC ACC TAC TAC AAC CCG 42

(2) INFORMATION FOR SEQ ID NO:39

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 36 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

GGG AGC TAC GGG GGC TAC TAC TAC GGT ATG GAC GTC 36

(2) INFORMATION FOR SEQ ID NO:40

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

GAT GCA TTG CCA AAG CAA TAT GCT TAT 27

(2) INFORMATION FOR SEQ ID NO:41

- 61 -

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 12 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

AAA GAC AGT GAG 12

(2) INFORMATION FOR SEQ ID NO:42

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 33 base pairs

(B) TYPE: nucleic acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(G) CELL TYPE: Hybridoma producing human antibody

1-3-1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

CAA TCA GCA GAC AGC AGT GGT ACT TAT GAG GTA

33